

# Imagen® SYBR Green qPCR Premix

Cat. NO IPQ

## Description

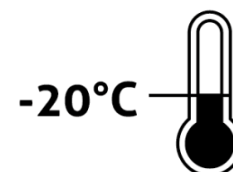
**Imagen® SYBR Green qPCR Premix** is a ready-to-use, 2X concentrated reagent including Hotstart Taq, SYBR Green I and ROX Reference dye, specially designed for real-time PCR with intercalator method. The ROX is optimal for instruments from Applied Biosystems (models 7000, 7300, 7700, 7900HT, StepOne™, and StepOnePlus™, but not Applied Biosystems 7500 Real Time PCR Systems, Smart Cycler® or LightCycler® real time instruments.) The ROX Reference dye does not participate in the PCR amplification.

## Contents

The **Imagen® SYBR Green qPCR Premix** is supplied as a ready-to-use 2x reaction mix. The formulation contains Hotstart Taq DNA polymerase, MgCl<sub>2</sub>, dNTPs, ROX Reference Dye, reaction enhancers, and stabilizers.

## Storage

- ✓ -20 °C
- ✓ Protected from light
- ✓ Avoid repeated freezing and thawing



## Reaction Mix Thawing and Handling

**Imagen® SYBR Green qPCR Premix** is delivered in a 2x ready-to-use format. To use the mix, thaw the vial on ice to 4 °C.

Please completely mix the vial and briefly centrifuge to ensure all components are at the bottom of the tube. Store on ice protected from light until ready to use. If using automated liquid handling, let sit at ambient temperature for 10 min to further reduce the viscosity.

## Application

- ✓ Detection and Quantification of DNA targets.
- ✓ High Throughput Applications.

## Prepare the qPCR Reaction Mix

1. Mix the **Imagen® SYBR Green qPCR Premix** thoroughly but gently until it's completely homogenous.
2. Prepare the qPCR Reaction Mix for the number of reactions required as shown in table below and plus 10% overage.

| Reagent                        | Volume (ul) | Final conc.    |
|--------------------------------|-------------|----------------|
| Imagen® SYBR Green qPCR Premix | 12.5        | 1x             |
| Forward Primer(10 uM)          | 0.75        | 300 - 600 nM   |
| Reverse Primer(10 uM)          | 0.75        | 300 - 600 nM   |
| DNA Template                   | 2           | 100 ng - 10 pg |
| Nuclease-free water            | 9           | -              |
| <b>Final volume</b>            | <b>25</b>   | <b>-</b>       |

3. Vortex the tube to mix the contents thoroughly, then centrifuge briefly to collect the contents at the bottom of the tube. (\*Use good pipetting practice to ensure assay precision and accuracy of dispensing.)

4. Add DNA (and nuclease-free water, if needed) to the PCR tubes or wells containing the reaction mix, seal tubes or wells with flat caps or optically transparent film, and gently vortex to ensure thorough mixing of the reaction components.
5. Program the thermal cycling protocol on the real-time PCR instrument.

| Step                                                |                                      | Temp. °C             | Time                         | Cycles |
|-----------------------------------------------------|--------------------------------------|----------------------|------------------------------|--------|
| DNA polymerase activation and template denaturation |                                      | 95°C                 | 10 min                       | 1      |
| Amplification                                       | Template denaturation                | 95°C                 | 20 sec                       | 35-40  |
|                                                     | Annealing / Extension and plate read | 58 - 62°C            | 60 sec<br>© Data acquisition |        |
| Melt Curve                                          |                                      | 95°C<br>60°C<br>95°C | 20 sec<br>20 sec<br>15 sec   | 1      |

6. Load the PCR tubes or plates onto the real-time PCR instrument and start the qPCR run program.
7. When thermal cycling is complete, perform data according to the instructions in the instrument-specific software.